TEST REPORT

1. NO : CU25-01167E
2. Client
   - ○ Name : NEWFULLY co., ltd.
   - ○ Address : 201-1201, 589-8 Siheung-daero, Yeongdeungpo-gu, Seoul, Republic of Korea
3. Date of Test : 2025.05.07 ~ 2025.06.04
4. Use of Report : For reference only(R&D)
5. Test Sample : Weefree wet wipes[Expiration date:1 year]
6. Test Method
   (1) Specification and test method of the client
   (2) The regulation on the safety standards of cosmetics and others

Affirmation	Tested By Name : Kim Mijin	Technical Manager Name : Byung Rye Go

This report is not accredited by KOLAS and KS Q ISO/IEC 17025.
Our report apply only to the standards or procedures identified and to the sample(s) tested unless otherwise specified. The test results are not indicative of representative of the qualities of the lot from which the sample was taken or of apparently identical or similar products. The results of using only a portion of this report cannot be guaranteed. The authenticity of this test report can be checked on KCL website(www.kcl.re.kr).

2025.06.04

Korea Conformity Laboratories

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7. Test range

Test Item			Test method
Appearance(Including external appearance)			(1)
pH			(1)
Weight change rate			(1)
Limit of microorganisms	Total aerobic viable cell count	Bacteria count	(2)
		Fungus count	
	Specified bacteria	*Escherichia coli*	
		Pseudomonas aeruginosa	
		Staphylococcus aureus	

8. Test condition of stability test and assessment method

8.1. Test condition of stability test

8.1.1. Storage condition

8.1.1.1. Accelerated storage condition : 40 ℃

8.1.1.2. Stress storage condition : -15 ℃

8.1.2. Assessment period : Initial(D_0), After 10(D_{10}) days of conditioning

8.2. Test method

8.2.1. Appearance

8.2.1.1. After the storage period described 6.1., visually observe changes in appearance. As a result of visual judgment, if it meets the criteria presented by the client without any changes, it is judged as 'conforms'.

8.2.2. pH

8.2.2.1. After the designated storage period has elapsed, take approximately 2 g of the sample and place it in a 100 mL beaker. Add 30 mL of water, heat the mixture in a water bath to dissolve the fatty substances, and stir to mix thoroughly. Then cool it in a refrigerator to solidify the fats and filter the mixture. If the fat and water layers are not clearly separated, use the mixture as is. However, in the case of clear liquid formulations depending on the product characteristics, the pH may be measured directly without the above procedure.

8.2.3. Weight change rate

8.2.3.1. After the designated storage period has elapsed, weigh the product and calculate the weight change rate using the following formula.

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$$Weight\ change\ rate(\%) = \frac{T_X - T_0}{T_0} \times 100$$

T_X : Measured weight at Day 10 (D_{10})

T_0 : Measured weight at Day 0 (D_0)

8.2.4. Limit of microorganisms

8.2.4.1. Total aerobic viable cell count test

8.2.4.1.1. Growth promotion of the media and suitability of the counting method in the presence of product

8.2.4.1.1.1. Negative Control

8.2.4.1.1.1.1. To verify testing conditions, a negative control is performed using the chosen diluent in place of the test preparation. There must be no growth of microorganisms.

8.2.4.1.1.2. Growth promotion of the media

8.2.4.1.1.2.1. Test each batch of medium prepared from dehydrated medium.

8.2.4.1.1.2.2. Use standardized stable suspensions of test strains. Use Buffered Sodium Chloride-Peptone Solution pH 7.0 to make test suspensions

8.2.4.1.1.2.3. Inoculate petri dish of tryptic soy agar (TSA) or sabouraud dextrose agar (SDA) with a small number (not more than 100 CFU) of the microorganisms indicated in the table below, using a separate petri dish of medium for each. Incubate according to the conditions described in the table below.

8.2.4.1.1.2.4. For a freshly prepared inoculum, growth of the microorganisms comparable to that previously obtained with a previously tested and approved batch of medium occurs.

Microorganism	Incubation condition
Escherichia coli (ATCC 8739)	TSA(Under aerobic condition) ≦ 100 CFU, (30 ~ 35) ℃, ≧ 48 h
Bacillus subtilis (ATCC 6633)	
Staphylococcus aureus (ATCC 6538)	
Candida albicans (ATCC 10231)	SDA(Under aerobic condition) ≦ 100 CFU, (20 ~ 25) ℃, ≧ 5 days

8.2.4.1.1.3. Suitability of the counting method in the presence of product

8.2.4.1.1.3.1. Preparation of the sample

8.2.4.1.1.3.1.1. Carry out the determination under conditions designed to avoid extrinsic microbial contamination of the product to be examined.

8.2.4.1.1.3.1.2. Add 1 g of inactivator polysorbate 80 to 1 g of a sample, make it sufficiently homogenized, and add 8 mL of D/E Neutralizing Broth to prepare a 10-fold dilution solution.

8.2.4.1.1.3.2. Medium

8.2.4.1.1.3.2.1. For the Bacteria count test, tryptic soy agar (TSA) is used, and for the Fungus

count test, sabouraud dextrose agar (SDA) is used.

8.2.4.1.1.3.3. Performing the test

8.2.4.1.1.3.3.1. Prepare the test sample according to 8.2.4.1.1.3.1. and prepare the control without the sample. Inoculate the sample and the control with the strain used in 8.2.4.1.1.2.4. to not exceed 100 CFU and incubate.

8.2.4.1.1.3.3.2. After incubation, check the nature of the colonies and compare the count with a control (without the sample). The number of microorganisms recovered is compared the number of microorganisms recovered from the control preparation. If the count is less than 50% of the control, modify the procedure (diluents, neutralization agents or combination of both).

Interfering substance in cosmetics	Chemical compounds able to neutralize preservative's antimicrobial activity
Phenolic compounds : Parabens, phenoxyethanol, phenylethanol, etc. Anilides	Lecithin, Polysorbate 80, Ethylene oxide condensate of fatty alcohol, Non-ionic surfactants
Quaternary ammonium compounds, Cationic surfactants	Lecithin, saponin, polysorbate 80, sodium dodecyl sulfate, Ethylene oxide condensate of fatty alcohol
Aldehydes, Formaldehyde-release agents	Glycine, histidine
Oxidizing compounds	Sodium thiosulfate
Isothiazolinones, imidazoles	Lecithin, saponin, Amines, sulfates, mercaptans, sodium bisulfite, sodium thioglycollate
Biguanides	Lecithin, saponin, polysorbate 80
Metallic salts (Cu, Zn, Hg), Organo-mercuric compounds	Sodium bisulfate, L-cysteine, Sulfhydryl compounds, thioglycollic acid

8.2.4.1.2. Examination of the Product

8.2.4.1.2.1. Preparation of the sample

8.2.4.1.2.1.1. Carry out the determination under conditions designed to avoid extrinsic microbial contamination of the product to be examined.

8.2.4.1.2.1.2. Add 1 g of inactivator polysorbate 80 to 1 g of a sample, make it sufficiently homogenized, and add 8 mL of D/E Neutralizing Broth to prepare a 10-fold dilution solution.

8.2.4.1.2.2. Medium

8.2.4.1.2.2.1. For the Bacteria count test, tryptic soy agar (TSA) is used, and for the Fungus count test, sabouraud dextrose agar (SDA) is used.

8.2.4.1.2.3. Performing the test

8.2.4.1.2.3.1. Bacteria count test (Pour-Plate Method)

8.2.4.1.2.3.1.1. For petri dishes 9 cm in diameter, add to the dish 1 mL of the sample prepared

and 15 to 20 mL of tryptic soy agar (TSA) media maintained at not more than 45 ℃.

8.2.4.1.2.3.1.2. Use at least two dishes per sample and incubate at (30 ~ 35) ℃ for at least 48 hours. Use petri dish with the maximum number of colonies, but measure the bacteria count with 300 or less colonies per petri dish as the maximum.

8.2.4.1.2.3.2. Fungus count test (Pour-Plate Method)

8.2.4.1.2.3.2.1. For petri dishes 9 cm in diameter, add to the dish 1 mL of the sample prepared and 15 to 20 mL of sabouraud dextrose agar (SDA) media maintained at not more than 45 ℃.

8.2.4.1.2.3.2.2. Use at least two dishes per sample and incubate at (20 ~ 25) ℃ for at least 5 days. Measure the fungus count by counting petri dish with less than 100 colonies.

8.2.4.1.3. Expression of results

8.2.4.1.3.1. Determine the dilution factor of the test solution based on the results of the suitability of the counting method in the presence of product. If multiple dilution levels are used, select the plate showing the highest number of colonies.

8.2.4.1.3.2. If the average number of colonies on the plate inoculated with 1 mL of the test solution is less than 1, express the result as: < [dilution factor] CFU/g

8.2.4.1.3.3. If the average number of colonies on the plate inoculated with 1 mL of the test solution is less than 1, express the result as: < [dilution factor] CFU/g

8.2.4.2. Test for specified bacteria

8.2.4.2.1. Growth promotion of the media and suitability of the counting method in the presence of product

8.2.4.2.1.1. Negative Control

8.2.4.2.1.1.1. To verify testing conditions, a negative control is performed using the chosen diluent in place of the test preparation. There must be no growth of microorganisms.

8.2.4.2.1.2. Growth promotion of the media

8.2.4.2.1.2.1. Test each batch of medium prepared from dehydrated medium.

8.2.4.2.1.2.2. Use standardized stable suspensions of test strains. Use Buffered Sodium Chloride-Peptone Solution pH 7.0 to make test suspensions.

8.2.4.2.1.2.3. Each strain shall be individually inoculated into the medium at 100 CFU and incubated under its respective conditions.

8.2.4.2.1.2.4. For a freshly prepared inoculum, growth of the microorganisms comparable(positive) to that previously obtained with a previously tested and approved batch of medium occurs.

8.2.4.2.1.3. Suitability of the counting method in the presence of product

8.2.4.2.4.3.1. Each test strain shall be inoculated at 100 CFU into the test and control solutions prepared with and without the sample, according to the specified microbial limit test

method, and incubated as directed. Each strain shall yield a positive result.

8.2.4.2.4.3.2. If microbial growth is inhibited (negative result) due to antimicrobial activity of preservatives or other substances in the sample, dilution or neutralizers(8.2.4.1.1.3.3.) may be used to ensure the validity of the results.

Microorganism	Incubation condition
Escherichia coli (ATCC 8739)	LB, (30 ~ 35) ℃, (24 ~ 72) h
	MAC agar, (30 ~ 35) ℃, (18 ~ 24) h
Pseudomonas aeruginosa (ATCC 9027)	TSB, (30 ~ 35) ℃, (24 ~ 48) h
	CET agar, (30 ~ 35) ℃, (24 ~ 48) h
Staphylococcus aureus (ATCC 6538)	TSB, (30 ~ 35) ℃, (24 ~ 48) h
	BP agar, (30 ~ 35) ℃, 24 h

8.2.4.2.2. Examination of the Product

8.2.4.2.2.1. Test for *Escherichia coli*

8.2.4.2.2.1.1. Preparation of the sample

8.2.4.2.2.1.1.1. Carry out the determination under conditions designed to avoid extrinsic microbial contamination of the product to be examined.

8.2.4.2.2.1.1.2. Prepare the test solution by diluting 1 g of the sample with 10 mL of lactose broth (1:10 dilution), and incubate at (30 ~ 35) ℃ for (24 ~ 72) hours.

8.2.4.2.2.1.2. Selection and subculture

8.2.4.2.2.1.2.1. Subculture on a plate of MacConkey Agar, and incubate at (30 ~ 35) ℃ for (18 ~ 24) hours.

8.2.4.2.2.1.3. Judgment

8.2.4.2.2.1.3.1. If no reddish-brown Gram-negative colonies with a red precipitate ring surrounding them are detected, the sample is considered negative for *Escherichia coli*.

8.2.4.2.2.1.3.2. If colonies of reddish-brown Gram-negative bacteria with a surrounding red precipitate ring are observed, streak each colony onto eosin methylene blue (EMB) agar and incubate at (30 ~ 35) ℃ for (18 ~ 24) hours.

8.2.4.2.2.1.3.3. If colonies showing a metallic sheen or dark blue-black coloration under transmitted light are observed on EMB agar, transfer a single colony using a platinum loop into lactose broth containing a fermentation tube, and incubate in a water bath at (44.3 ~ 44.7) ℃ for (22 ~ 26) hours.

8.2.4.2.2.1.3.4. If gas production is observed, suspect the presence of *Escherichia coli* and confirm by identification testing.

8.2.4.2.2.2. Test for *Pseudomonas aeruginosa*

8.2.4.2.2.2.1. Preparation of the sample

8.2.4.2.2.2.1.1. Carry out the determination under conditions designed to avoid extrinsic microbial contamination of the product to be examined.

8.2.4.2.2.2.1.2. Prepare the test solution by diluting 1 g of the sample with 10 mL of tryptic soy broth (1:10 dilution), and incubate at (30 ~ 35) ℃ for (24 ~ 48) hours.

8.2.4.2.2.2.2. Selection and subculture

8.2.4.2.2.2.2.1. Subculture on a plate of Cetrimide Agar, and incubate at (30 ~ 35) ℃ for (24 ~ 48) hours.

8.2.4.2.2.2.3. Judgment

8.2.4.2.2.2.3.1. If no microbial growth is observed, the sample is considered negative for *Pseudomonas aeruginosa*.

8.2.4.2.2.2.3.2. If colonies of Gram-negative rods exhibiting green fluorescence are observed, the enrichment culture is streaked onto Pseudomonas agar base media P and F, and incubated at (30 ~ 35) ℃ for (24 ~ 72) hours.

8.2.4.2.2.2.3.3. Colonies on Pseudomonas agar F (for fluorescein detection) are examined under UV light for yellow fluorescence, and colonies on Pseudomonas agar P (for pyocyanin detection) are examined under UV light for blue pigmentation.

8.2.4.2.2.2.3.4. If such colonies are observed, an oxidase test is performed. A positive oxidase reaction is indicated by the appearance of a purple color within (5 ~ 10) seconds. If no color change is observed within 10 seconds, the result is considered negative for *Pseudomonas aeruginosa*. If the oxidase reaction is positive, the presence of *Pseudomonas aeruginosa* is suspected, and further identification testing shall be performed for confirmation.

8.2.4.2.2.3. Test for *Staphylococcus aureus*

8.2.4.2.2.3.1. Preparation of the sample

8.2.4.2.2.3.1.1. Carry out the determination under conditions designed to avoid extrinsic microbial contamination of the product to be examined.

8.2.4.2.2.3.1.2. Prepare the test solution by diluting 1 g of the sample with 10 mL of tryptic soy broth (1:10 dilution), and incubate at (30 ~ 35) ℃ for (24 ~ 48) hours.

8.2.4.2.2.3.2. Selection and subculture

8.2.4.2.2.3.2.1. Subculture on a plate of Baird Parker Agar, and incubate at (30 ~ 35) ℃ for 24 hours.

8.2.4.2.2.3.3. Judgment

8.2.4.2.2.3.3.1. If the colony appears black with a surrounding yellow-clear zone and is identified as Gram-positive by Gram staining, a coagulase test shall be performed.

8.2.4.2.2.3.3.2. If the coagulase test is negative, the sample is considered negative for *Staphylococcus aureus*. If positive, the presence of *Staphylococcus aureus* is suspected and shall be confirmed by identification testing.

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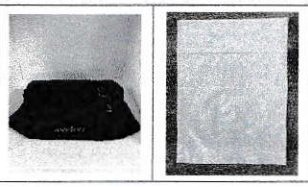
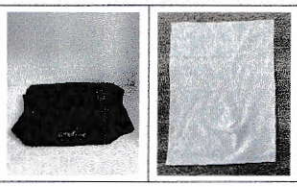
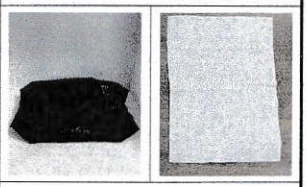
9. Test results

9.1. Appearance

Test item	Specification	D_0	D_{10}	
			40 ℃	-15 ℃
Appearance	Wet tissue impregnated with liquid formulation			
		Conforms	Conforms	Conforms

9.2. pH

Test item	Specification	D_0	D_{10}	
			40 ℃	-15 ℃
pH	-	5.2	5.0	5.0

9.3. Weight change rate

Test item	D_0		D_{10}	
	40 ℃	-15 ℃	40 ℃	-15 ℃
Weight(g)	940.95	942.65	934.36	942.91
Weight change rate(%)	-	-	-0.70	0.03

9.4. Initial microbial limit test

9.4.1. Total aerobic viable cell count

9.4.1.1. Negative Control

Media	Growth of microorganisms
Tryptic soy agar (TSA)	No Growth
Sabouraud dextrose agar (SDA)	No Growth

9.4.1.2. Growth promotion of the media

Media	Microorganisms	Inoculation	Incubation condition	Judgment
Tryptic soy agar (TSA)	*Escherichia coli* (ATCC 8739)	100 CFU or less	(30 ~ 35) ℃, 48 h	PASS

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	Bacillus subtilis (ATCC 6633)	100 CFU or less	(30 ~ 35) ℃, 48 h	PASS
	Staphylococcus aureus (ATCC 6538)	100 CFU or less	(30 ~ 35) ℃, 48 h	PASS
Sabouraud dextrose agar (SDA)	*Candida albicans* (ATCC 10231)	100 CFU or less	(20 ~ 25) ℃, 5 days	PASS

9.4.1.3. Suitability of the counting method in the presence of product

9.4.1.3.1. The method suitability of bacterial count test was confirmed under 10-fold dilution conditions using D/E Neutralizing Broth as the diluent and polysorbate 80 as the dispersing agent.

9.4.1.3.2. The method suitability of the fungal count test was confirmed under 10-fold dilution conditions using D/E Neutralizing Broth as the diluent and polysorbate 80 as the dispersing agent.

Microorganisms	Inoculation	Incubation condition	Diluent/ Dispersing agent/ Dilution factor	Judgment
Escherichia coli (ATCC 8739)	100 CFU or less	TSA, (30 ~ 35) ℃, 48 h	D/E Neutralizing Broth/ polysorbate 80/ 10-fold	PASS
Bacillus subtilis (ATCC 6633)	100 CFU or less			
Staphylococcus aureus (ATCC 6538)	100 CFU or less			
Candida albicans (ATCC 10231)	100 CFU or less	SDA, (20 ~ 25) ℃, 5 days	D/E Neutralizing Broth/ polysorbate 80/ 10-fold	PASS

9.4.1.4. Examination of the Product

Test item		Specification (CFU/g)	Test results (CFU/g)	Judgment
Total aerobic viable cell count	Bacteria count	100 of less	< 10	PASS
	Fungus count	100 of less	< 10	PASS

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9.4.2. Specified bacteria test

9.4.2.1. *Escherichia coli*

9.4.2.1.1. Negative Control

Media	Growth of microorganisms
Lactose Broth (LB)	No Growth
MacConkey Agar (MAC agar)	No Growth

9.4.2.1.2. Growth promotion of the media

Media	Microorganisms	Incubation condition	Judgment
LB	*Escherichia coli* (ATCC 8739)	(30 ~ 35) ℃, (24 ~ 72) h	PASS
MAC agar	*Escherichia coli* (ATCC 8739)	(20 ~ 25) ℃, 5 days	PASS

9.4.2.1.3. Suitability of the counting method in the presence of product

Microorganisms	Sample preparation and pre-incubation	Selection and subculture	Judgment
Escherichia coli (ATCC 8739)	LB(10-fold) (30 ~ 35) ℃, (24 ~ 72) h	MAC agar (30 ~ 35) ℃, (18 ~ 24) h	PASS

9.4.2.1.4. Examination of the Product

Sample preparation and pre-incubation	Selection and subculture	Test result	Judgment
LB(10-fold) (30 ~ 35) ℃, (24 ~ 72) h	MAC agar (30 ~ 35) ℃, (18 ~ 24) h	Negative	PASS

9.4.2.2. *Pseudomonas aeruginosa*

9.4.2.2.1. Negative Control

Media	Growth of microorganisms
Tryptic Soy Broth(TSB)	No Growth
Cetrimide Agar(CET agar)	No Growth

9.4.2.2.2. Growth promotion of the media

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Media	Microorganisms	Incubation condition	Judgment
TSB	*Pseudomonas aeruginosa* (ATCC 9027)	(30 ~ 35) ℃, (24 ~ 48) h	PASS
CET agar	*Pseudomonas aeruginosa* (ATCC 9027)	(30 ~ 35) ℃, (24 ~ 48) h	PASS

9.4.2.2.3. Suitability of the counting method in the presence of product

Microorganisms	Sample preparation and pre-incubation	Selection and subculture	Judgment
Pseudomonas aeruginosa (ATCC 9027)	TSB(10-fold) (30 ~ 35) ℃, (24 ~ 48) h	CET agar (30 ~ 35) ℃, (24 ~ 48) h	PASS

9.4.2.2.4. Examination of the Product

Sample preparation and pre-incubation	Selection and subculture	Test result	Judgment
TSB(10-fold) (30 ~ 35) ℃, (24 ~ 48) h	CET agar (30 ~ 35) ℃, (24 ~ 48) h	Negative	PASS

9.4.2.3. *Staphylococcus aureus*

9.4.2.3.1. Negative Control

Media	Growth of microorganisms
Tryptic Soy Broth(TSB)	No Growth
Baird Parker Agar(BP agar)	No Growth

9.4.2.3.2. Growth promotion of the media

Media	Microorganisms	Incubation condition	Judgment
TSB	*Staphylococcus aureus* (ATCC 6538)	(30 ~ 35) ℃, (24 ~ 48) h	PASS
BP agar	*Staphylococcus aureus* (ATCC 6538)	(30 ~ 35) ℃, (24 ~ 48) h	PASS

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9.4.2.3.3. Suitability of the counting method in the presence of product

Sample preparation and pre-incubation	Selection and subculture	Test result	Judgment
Staphylococcus aureus (ATCC 6538)	TSB(10-fold) (30 ~ 35) ℃, (24 ~ 48) h	BP agar (30 ~ 35) ℃, (24 ~ 48) 24 h	PASS

9.4.2.3.4. Examination of the Product

Sample preparation and pre-incubation	Selection and subculture	Test result	Judgment
TSB(10-fold)) (30 ~ 35) ℃, (24 ~ 48) h	BP agar (30 ~ 35) ℃, 24 h	Negative	PASS

9.5. After 10(D_{10}) days of conditioning microbial limit test

9.5.1. Total aerobic viable cell count

9.5.1.1. Negative Control

Media	Growth of microorganisms
Tryptic soy agar (TSA)	No Growth
Sabouraud dextrose agar (SDA)	No Growth

9.5.1.2. Growth promotion of the media

Media	Microorganisms	Inoculation	Incubation condition	Judgment
Tryptic soy agar (TSA)	*Escherichia coli* (ATCC 8739)	100 CFU or less	(30 ~ 35) ℃, 48 h	PASS
	Bacillus subtilis (ATCC 6633)	100 CFU or less	(30 ~ 35) ℃, 48 h	PASS
	Staphylococcus aureus (ATCC 6538)	100 CFU or less	(30 ~ 35) ℃, 48 h	PASS
Sabouraud dextrose agar (SDA)	*Candida albicans* (ATCC 10231)	100 CFU or less	(20 ~ 25) ℃, 5 days	PASS

9.5.1.3. Suitability of the counting method in the presence of product

9.5.1.3.1. The method suitability of bacterial count test was confirmed under 10-fold dilution conditions using D/E Neutralizing Broth as the diluent and polysorbate 80 as the dispersing agent.

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9.5.1.3.2. The method suitability of the fungal count test was confirmed under 10-fold dilution conditions using D/E Neutralizing Broth as the diluent and polysorbate 80 as the dispersing agent.

Microorganisms	Inoculation	Incubation condition	Diluent/ Dispersing agent/ Dilution factor	Judgment
Escherichia coli (ATCC 8739)	100 CFU or less	TSA, (30 ~ 35) ℃, 48 h	D/E Neutralizing Broth/ polysorbate 80/ 10-fold	PASS
Bacillus subtilis (ATCC 6633)	100 CFU or less			
Staphylococcus aureus (ATCC 6538)	100 CFU or less			
Candida albicans (ATCC 10231)	100 CFU or less	SDA, (20 ~ 25) ℃, 5 days	D/E Neutralizing Broth/ polysorbate 80/ 10-fold	PASS

9.5.1.4. Examination of the Product

Test item		Specification (CFU/g)	Test results (CFU/g)	Judgment
Total aerobic viable cell count	Bacteria count	100 of less	< 10	PASS
	Fungus count	100 of less	< 10	PASS

9.5.2. Specified bacteria test

9.5.2.1. *Escherichia coli*

9.5.2.1.1. Negative Control

Media	Growth of microorganisms
Lactose Broth (LB)	No Growth
MacConkey Agar (MAC agar)	No Growth

9.5.2.1.2. Negative Control

Media	Microorganisms	Incubation condition	Judgment
LB	*Escherichia coli* (ATCC 8739)	(30 ~ 35) ℃, (24 ~ 72) h	PASS

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MAC agar	*Escherichia coli* (ATCC 8739)	(20 ~ 25) ℃, 5 days	PASS

9.5.2.1.3. Suitability of the counting method in the presence of product

Microorganisms	Sample preparation and pre-incubation	Selection and subculture	Judgment
Escherichia coli (ATCC 8739)	LB(10-fold) (30 ~ 35) ℃, (24 ~ 72) h	MAC agar (30 ~ 35) ℃, (18 ~ 24) h	PASS

9.5.2.1.4. Examination of the Product

Sample preparation and pre-incubation	Selection and subculture	Test result	Judgment
LB(10-fold) (30 ~ 35) ℃, (24 ~ 72) h	MAC agar (30 ~ 35) ℃, (18 ~ 24) h	Negative	PASS

9.5.2.2. *Pseudomonas aeruginosa*

9.5.2.2.1. Negative Control

Media	Growth of microorganisms
Tryptic Soy Broth(TSB)	No Growth
Cetrimide Agar(CET agar)	No Growth

9.5.2.2.2. Growth promotion of the media

Media	Microorganisms	Incubation condition	Judgment
TSB	*Pseudomonas aeruginosa* (ATCC 9027)	(30 ~ 35) ℃, (24 ~ 48) h	PASS
CET agar	*Pseudomonas aeruginosa* (ATCC 9027)	(30 ~ 35) ℃, (24 ~ 48) h	PASS

9.5.2.2.3. Suitability of the counting method in the presence of product

Microorganisms	Sample preparation and pre-incubation	Selection and subculture	Judgment
Pseudomonas aeruginosa (ATCC 9027)	TSB(10-fold) (30 ~ 35) ℃, (24 ~ 48) h	CET agar (30 ~ 35) ℃, (24 ~ 48) h	PASS

TEST REPORT

NO : CU25-01167E

9.5.2.2.4. Examination of the Product

Sample preparation and pre-incubation	Selection and subculture	Test result	Judgment
TSB(10-fold) (30 ~ 35) ℃, (24 ~ 48) h	CET agar (30 ~ 35) ℃, (24 ~ 48) h	Negative	PASS

9.5.2.3. *Staphylococcus aureus*

9.5.2.3.1. Negative Control

Media	Growth of microorganisms
Tryptic Soy Broth(TSB)	No Growth
Baird Parker Agar(BP agar)	No Growth

9.5.2.3.2. Growth promotion of the media

Media	Microorganisms	Incubation condition	Judgment
TSB	*Staphylococcus aureus* (ATCC 6538)	(30 ~ 35) ℃, (24 ~ 48) h	PASS
BP agar	*Staphylococcus aureus* (ATCC 6538)	(30 ~ 35) ℃, (24 ~ 48) h	PASS

9.5.2.3.3. Suitability of the counting method in the presence of product

Sample preparation and pre-incubation	Selection and subculture	Test result	Judgment
Staphylococcus aureus (ATCC 6538)	TSB(10-fold) (30 ~ 35) ℃, (24 ~ 48) h	BP agar (30 ~ 35) ℃, (24 ~ 48) 24 h	PASS

9.5.2.3.4. Examination of the Product

Sample preparation and pre-incubation	Selection and subculture	Test result	Judgment
TSB(10-fold)) (30 ~ 35) ℃, (24 ~ 48) h	BP agar (30 ~ 35) ℃, 24 h	Negative	PASS

----- End of Report -----

[KCL]성적서 발송 (위프리 물티슈 Weefree wet wipes_안정성 시험[사용기한:1년])

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[KCL] 고객님께서 요청하신 성적서의 참조본을 메일로 발송해 드립니다.
- 본 메일은 발신전용 메일 이므로 회신을 통한 문의는 처리되지 않습니다.
- 담당자메일 : ksh928@kcl.re.kr

성적서 수령방법(직접,우편 또는 전자문서) 변경시 연락주시기 바랍니다.
다음의 [성적서 보기] 버튼을 클릭하여 성적서를 열람 및 인쇄하시기 바랍니다.

성적서 보기

참조용 성적서입니다.
- 연구원 전용 성적서 뷰어프로그램 수동설치 - [자료실]

[전자문서(디지털성적서) 시행 안내] *※시험분석신청서 작성 시*
- (출력물 선택) KCL 성적서 전용용지 인쇄 및 1회 출력
- **(전자문서 선택)** 성적서 **파일 다운로드** 및 **출력 무제한**

※ 중복선택은 불가하며, 전자문서(디지털성적서) 관련 상세내용은 [이용절차]를 참고해주세요.

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